



Developments in understanding transmission risk and efficacy of screening scenarios

Nico Lelie, Director Scientific Affairs

Vilnius, Lithuania, May 19 2010



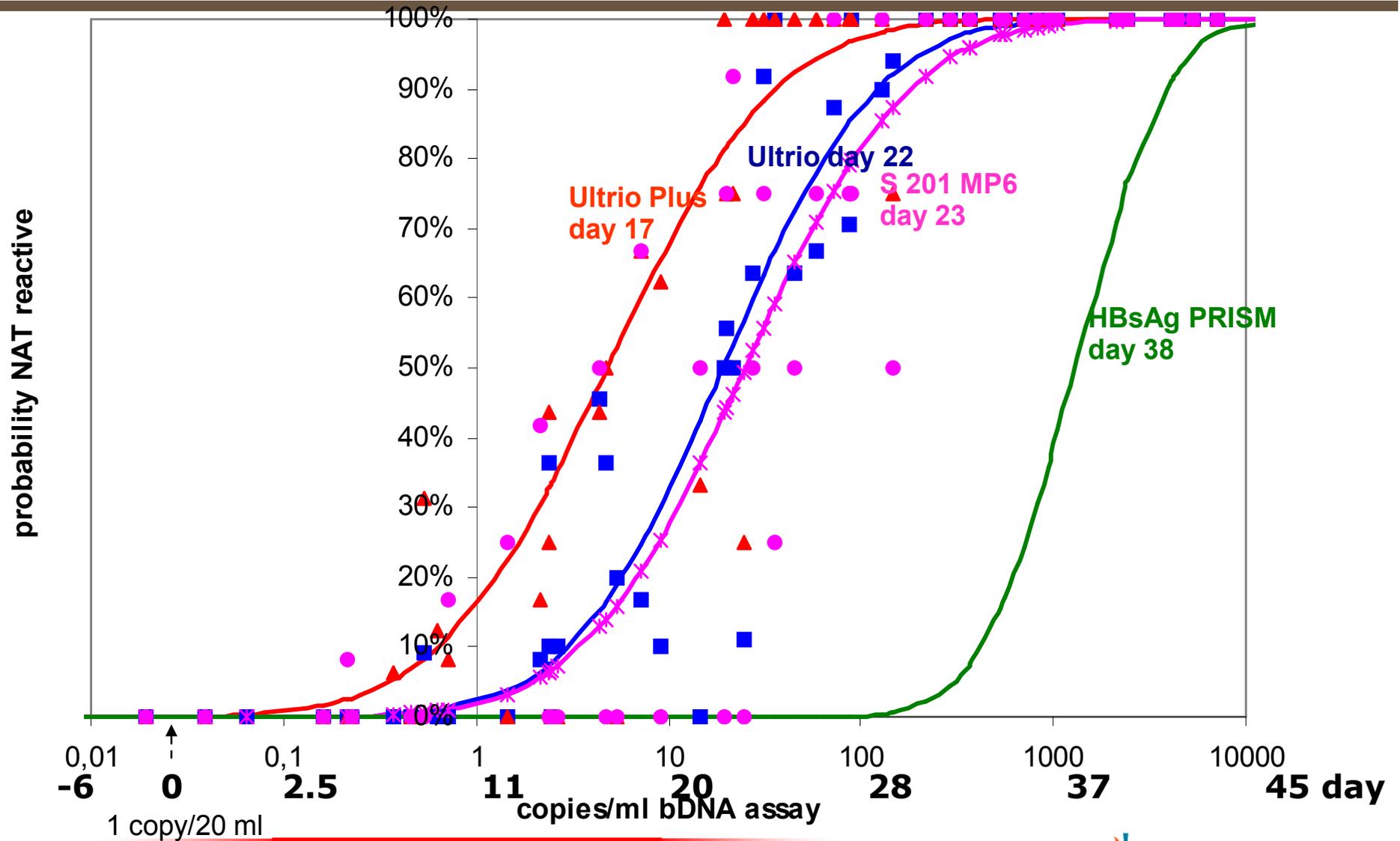
HBV genotype A2 panels and assays in EFS study¹

Panels	Assays
5 HBV genotype A seroconversion panels (Zeptomatrix 6254, 6289, 6292, 11006 and 11008) ¹	<ul style="list-style-type: none">■ Ultrio (5-18 replicates)■ Ultrio Plus (2-16 replicates)■ TaqScreen (4 replicates)■ bDNA 3.0 assay (single)■ PRISM HBsAg (2 replicates)■ Biorad HBsAg (2 replicates)
HBV-DNA genotype A standard dilution panel (PeliCheck, Sanquin-VQC) ^{1,2}	<ul style="list-style-type: none">■ Ultrio (24 replicates)■ Ultrio Plus (12 replicates)■ TaqScreen (12 replicates)

¹ EFS study (Assal et al, Transfusion 2009:49:289-300, Assal et al, Transfusion 2009:49:301-310)

² HBV standard was recalibrated in copies/ml by multiple bDNA 3.0 assays in DDL Diagnostic Laboratory (Voorburg, the Netherlands)

Probability of detection* of HBV by NAT methods and HBsAg in five seroconversion panels



Comparison of HBV NAT response on five seroconversion panels and on genotype A standard dilution series by probit analysis

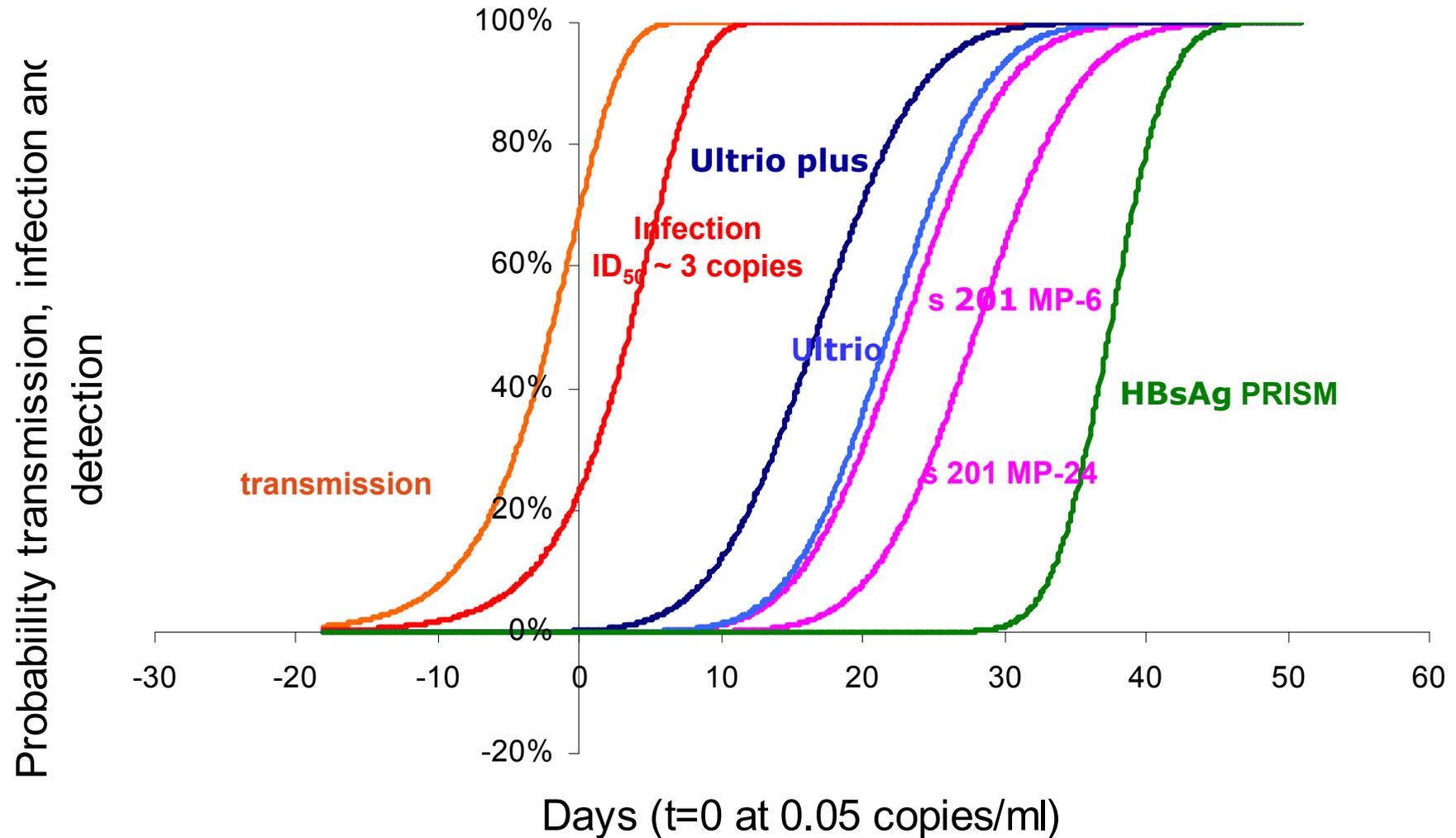
Assay	panels	Sam- ples	Repli- cates	50% LOD copies/ml	95% LOD copies/ml	Potency dilution to SC panels
Ultrio	5 SC	69	5-18	19 (15-26)	208 (140-347)	1.04 (0.62-1.74)
	dilution	10	24	19 (12-29)	200 (120-375)	
Ultrio Plus	5 SC	69	2-16	4.6 (3.4-6.3)	61 (38-110)	0.81 (0.41-1.56)
	dilution	10	12	5.7 (3.2-10)	75 (39-166)	
s 201 MP-6	5 SC	69	4	32 (20-49)	381 (206-913)	1.84 (0.63-8.85)
	dilution	10	12	17 (9.5-30)	207 (104-527)	

Analytical sensitivity* of NAT systems on seroconversion panels and HBV gt A dilution panel

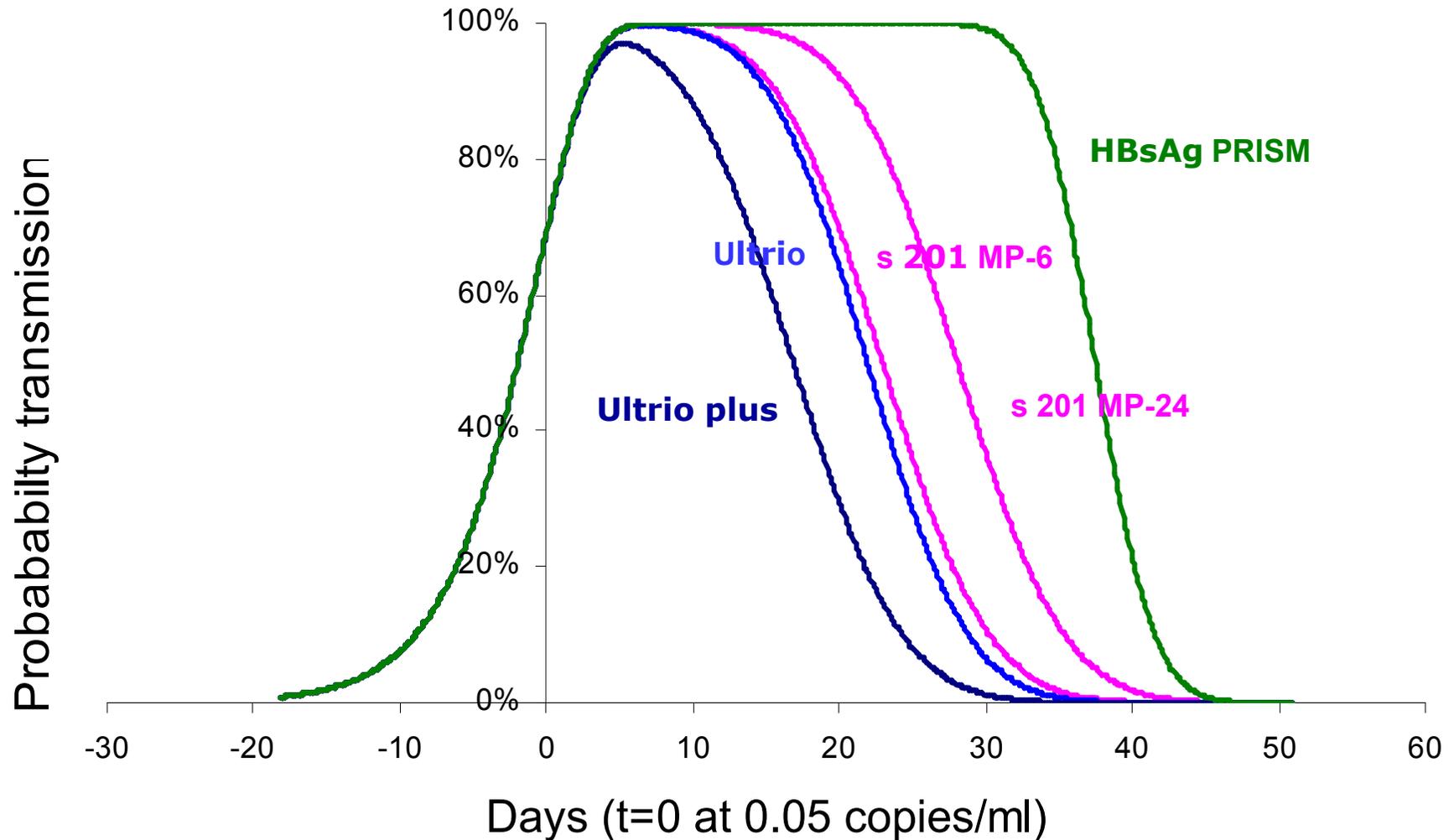
assay	50 % LOD (C.I.) copies/ml	95% LOD (C.I.) copies/ml
Ultrio ID	19 (16-24)	206 (146-319)
Ultrio plus ID	4.8 (3.7-6.4)	64 (41-113)
s 201 MP-6	25 (17-36)	316 (181-713)

*50 % and 95 % detection limits determined by probit analysis on seroconversion panels and genotype A standard dilution series taken together

Probability of transmission, infection and detection of HBV by NAT and HBsAg in infectious WP

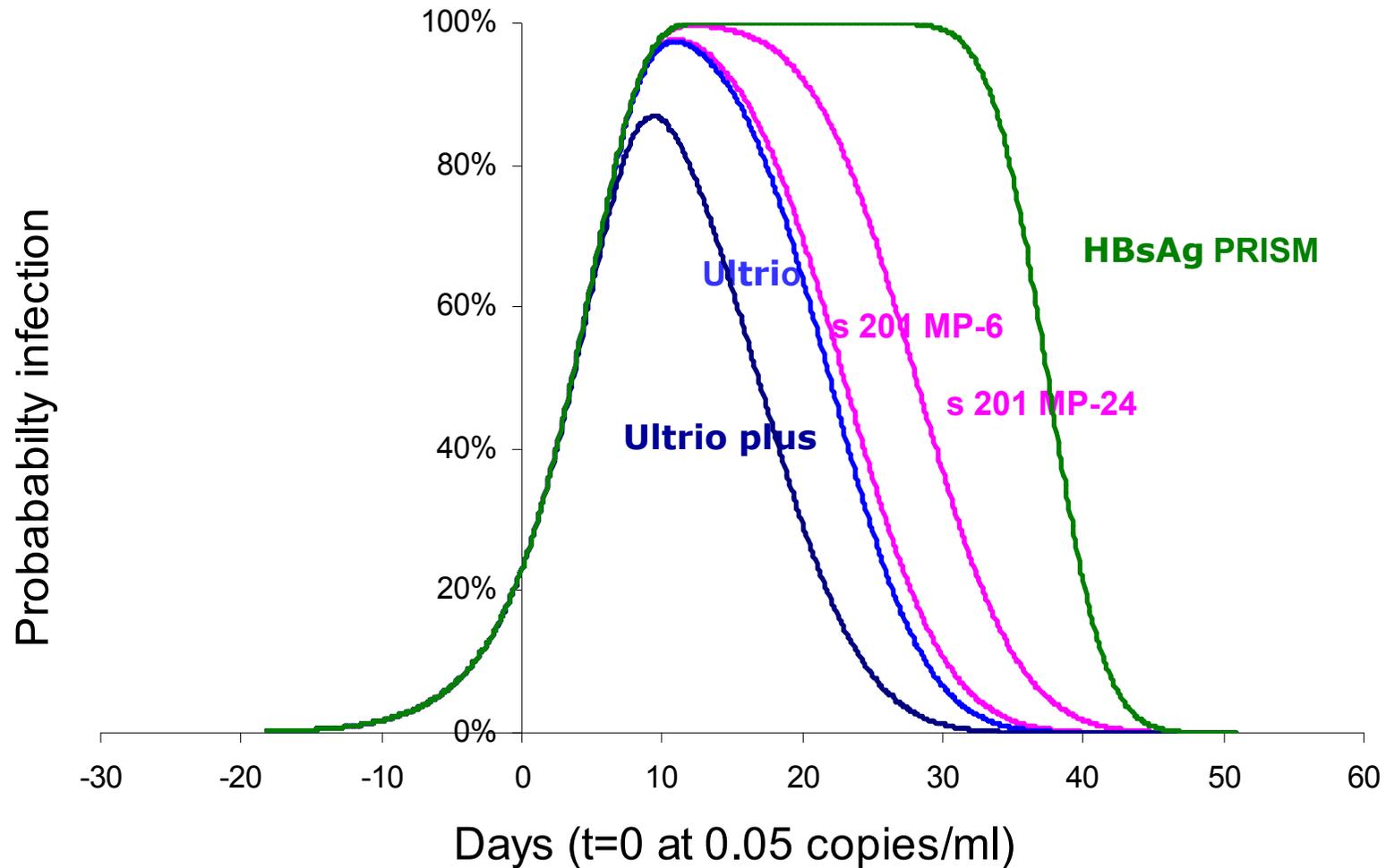


Risk of transmitting virions* with HBV-NAT and HBsAg testing



7 * Weusten et al. Transfusion, in press

Risk of HBV infection* with HBV-NAT and HBsAg testing (ID₅₀ ~ 3 copies)



* Weusten et al. Transfusion, in press

WP transmission risk day equivalents* with HBV-NAT and contribution of HBsAg testing

Assay	ID ₅₀ = 1 copy (virion)			ID ₅₀ = 3 copies (virions)		
	risk days	WP reduction by NAT	Extra WP reduction HBsAg	risk days	WP reduction by NAT	Extra WP reduction HBsAg
Ultrio Plus ID	19.6	20.6	0.0017	14.1	20.6	0.0017
Ultrio ID	24.7	15.5	0.013	19.1	15.6	0.013
s 201 MP-6	25.7	14.5	0.030	20.1	14.6	0.030
s 201 MP-24	30.8	9.4	0.22	25.3	9.4	0.22
PRISM	40.2			34.7		

* Weusten et al. Transfusion, in press

WP transmission risk day equivalents* with HBV-NAT and HBsAg testing, effect of ID₅₀

Assay	risk days				
	ID ₅₀ =1 copy	ID ₅₀ =3 <i>copies</i>	ID ₅₀ =10 copies	ID ₅₀ =30 copies	ID ₅₀ =100 copies
Ultrio Plus ID	19.6	14.1	9.6	6.1	3.3
Ultrio ID	24.7	19.1	14.4	10.4	6.5
s 201 MP-6	25.7	20.1	15.4	11.4	7.4
s 201 MP-24	30.8	25.3	20.5	16.4	12.0
PRISM	40.2	34.7	29.9	25.8	21.4

* Weusten et al. Transfusion, in press

New study protocol

- Multi-center analysis of the efficacy and cost effectiveness of HBV, HCV and HIV blood screening scenarios based on observed yield with serologic assays and individual donation nucleic acid amplification testing (ID NAT)
 - Sponsored by Novartis Diagnostics
 - Lead investigators: Steven Kleinman, Brian Custer, Michael Busch, Nico Lelie
 - Other investigators from multiple participating laboratories worldwide that use Ultrio ID NAT and chemiluminescent (or reasonably equivalent) serology assays

Primary study objectives

- Compare the efficacy of different possible NAT and serology screening scenarios in first time and repeat donors for HIV, HCV, and HBV infection
 - Calculate modelled residual risk based on observed ID NAT and serology yield in many countries
 - Efficacy is defined as percentage of transmission risk that is removed by a given testing strategy
- Compare the cost effectiveness of these scenarios
- Examine if different screening scenarios could be selected in low and high incidence/prevalence countries

Secondary study objectives

- Classify HIV, HCV, and HBV infections into various phases of infection and analyze these data by geographic region
 - South Africa, Egypt, Mediterranean, Eastern Europe, Nordic (North European), Southeast Asia, NZ
- Compare the ratio of cases detected by serology alone, ID NAT alone, and serology plus NAT in regions with differing infection incidence
- Extend the analysis to antigen/antibody combo assays and MP-NAT of various sizes and separately to lapsed donors

Input data required from each participating lab

- Number of donations tested:
 - classified by donor status: first time, repeat, and lapsed (> 1 year)
- Test yield data:
 - NAT only, serology only, and concordant NAT/serology classified by donor status
- More detailed data on positive donations
 - Pre-seroconversion or pre-NAT conversion interdonation interval
 - Data to validate true positivity and the phase of infection

Additional data for positive donations

- Differentiate true positives from false positives
 - Serology confirmatory testing procedures and detailed data (e.g., RIBA or Western blot band patterns)
 - Donor follow-up testing data
 - NAT data on an alternative sample (e.g., plasma bag)
 - Additional HBV assay results
- Classify stage of infection
 - Follow-up testing data
 - HIV: p24 Ag status, Western blot results if using combined Ag-Ab EIA
 - HBV: vaccination history, anti-HBc status, anti-HBc IgM status, anti-HBs titer in mIU/mL

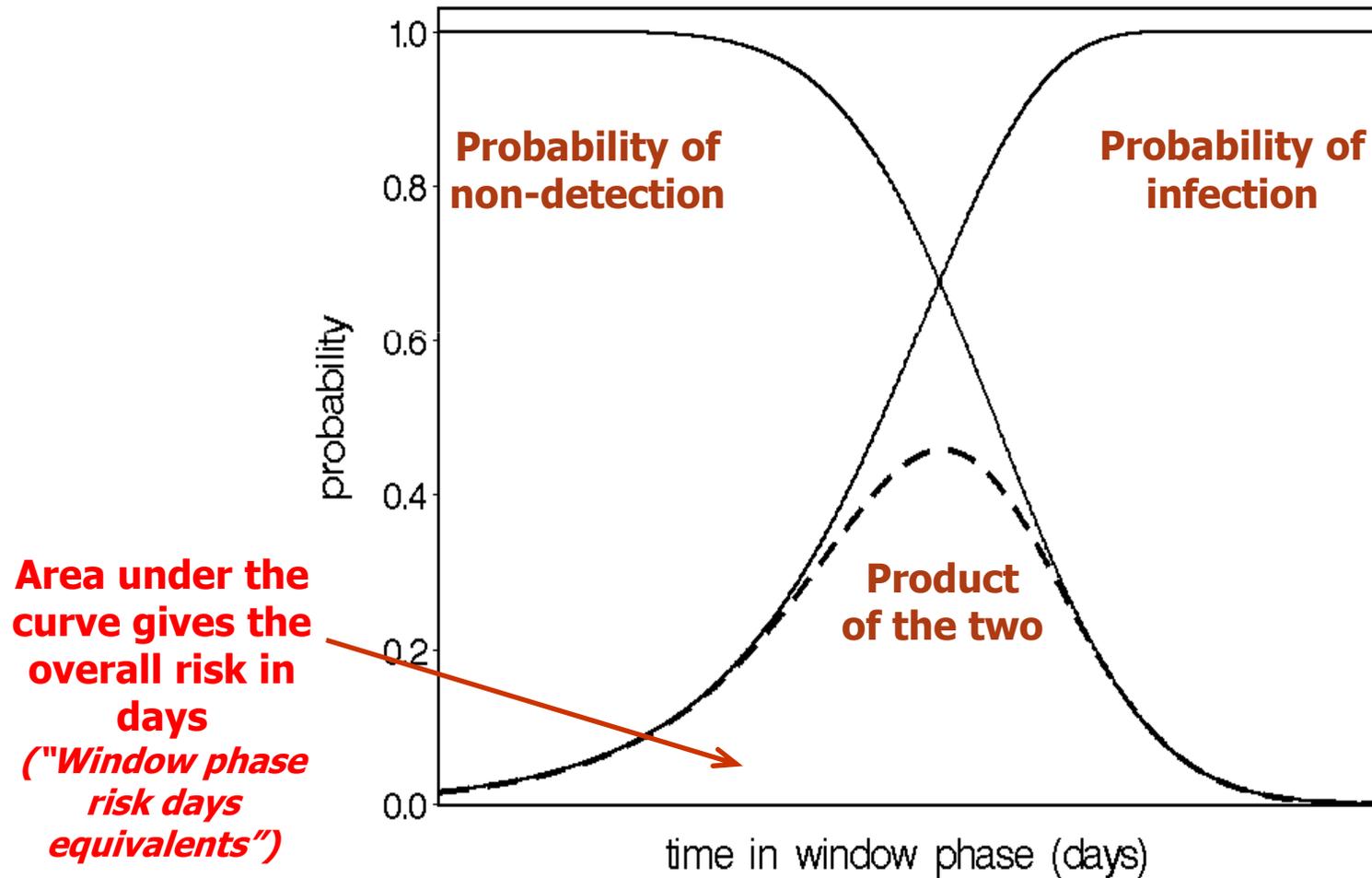
Data standardization issues

- Determining when data from different countries can/should be combined
- Definition of a “lapsed” donation
 - proposed to be a repeat donation with an interdonation interval of > one year
- Missing data
 - particularly impacts classifying the phases of HBV infection

Model for residual risk

- Calculate residual risk for packed RBC transfusion
- Use the statistical risk day equivalent model (Weusten et al) to calculate risk in repeat donors
 - Baseline scenario is ID NAT plus serology: this gives minimal residual risk
 - Evaluate residual risk in other testing scenarios: ID NAT only, serology only; MP NAT at various pool sizes
- Calculate risk in first time donors by using the ratio of ID NAT yield rate in first time to repeat donors as a conversion factor

Probability of infectivity during the window period (Weusten et al, Transfusion, in press)

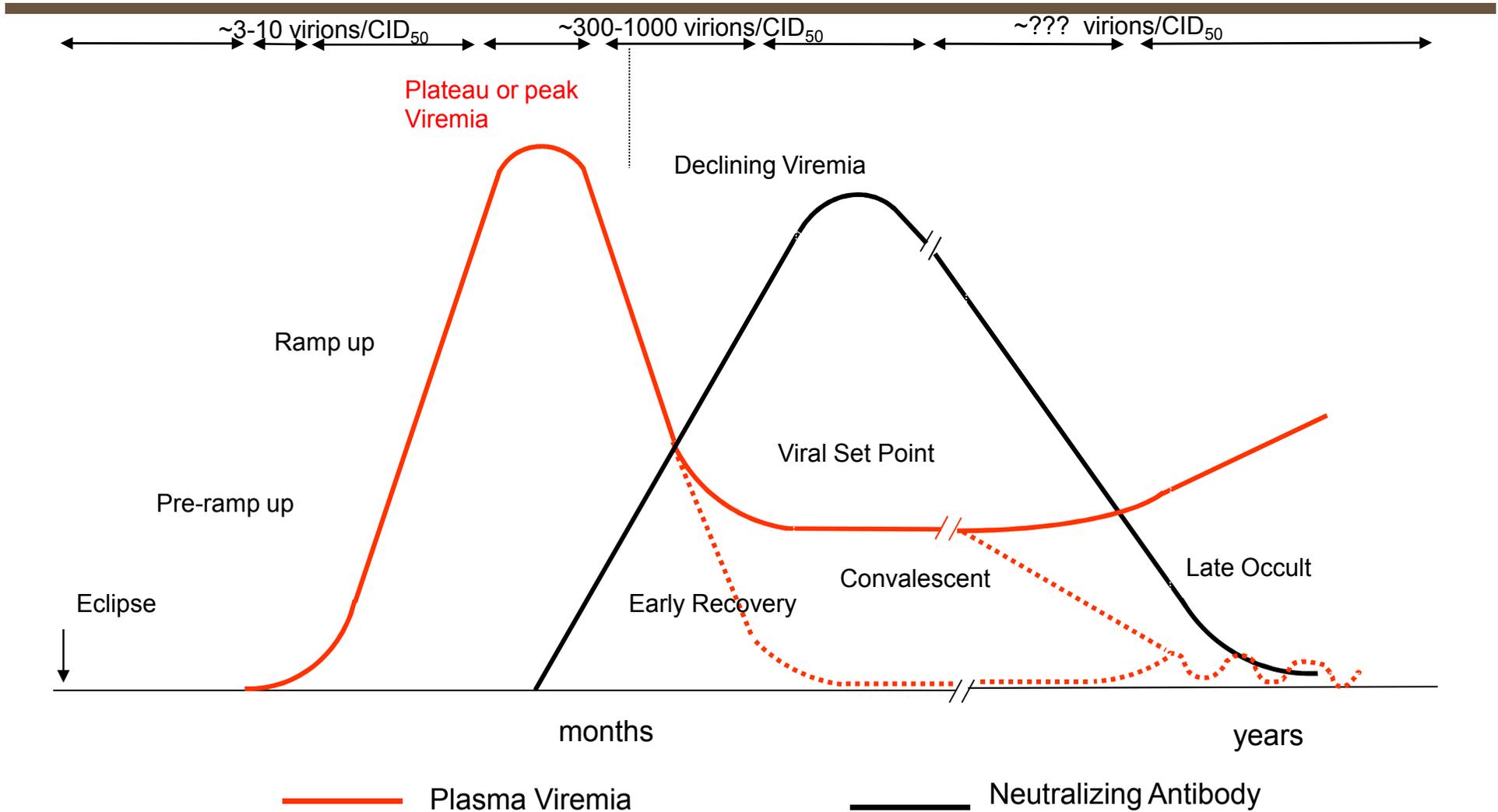


Standardization and input parameters for modelling

- Calibration of NAT detection limits and minimum infectious dose in virions or nucleic acid copies using bDNA 3.0 assay as reference method^{1,2}
- Determination of seroconversion point in antigen and combo assays during ramp up phase in copies/ml (bDNA assay)^{3,4,5}
- Random appearance of donors in WP³
- Half life of nucleic acid and antigen in clearance phase⁶
- Minimum infectious dose in different stages of infection^{7,8,9,10}

1). Van Drimmelen A.A.J. Vox Sang 96 Suppl1 ISBT abstract P075. 2). Weusten J et al, Transfusion, in press 3). Vermeulen M et al, Vox Sang ISBT abstract (in press). 4). LaPerche et al, to be published. 5). Van Drimmelen et al. Vox Sang ISBT abstract (in press) 6). Yoshikawa A et al, Vox Sang 2005;88:77-86 7) Komiya K et al. Transfusion 2008;48:286-8 8). Katayama K et al, Intervirology, 2004, 47, 57 9). Tabuchi A et al. J Med Virol 2008;80:2064-8 10) Ma ZM et al, J of Virology, 2009, 83:3288-97

Schematic of phases of infection



Assumptions for minimum infectious dose (ID_{50}) in different infection stages

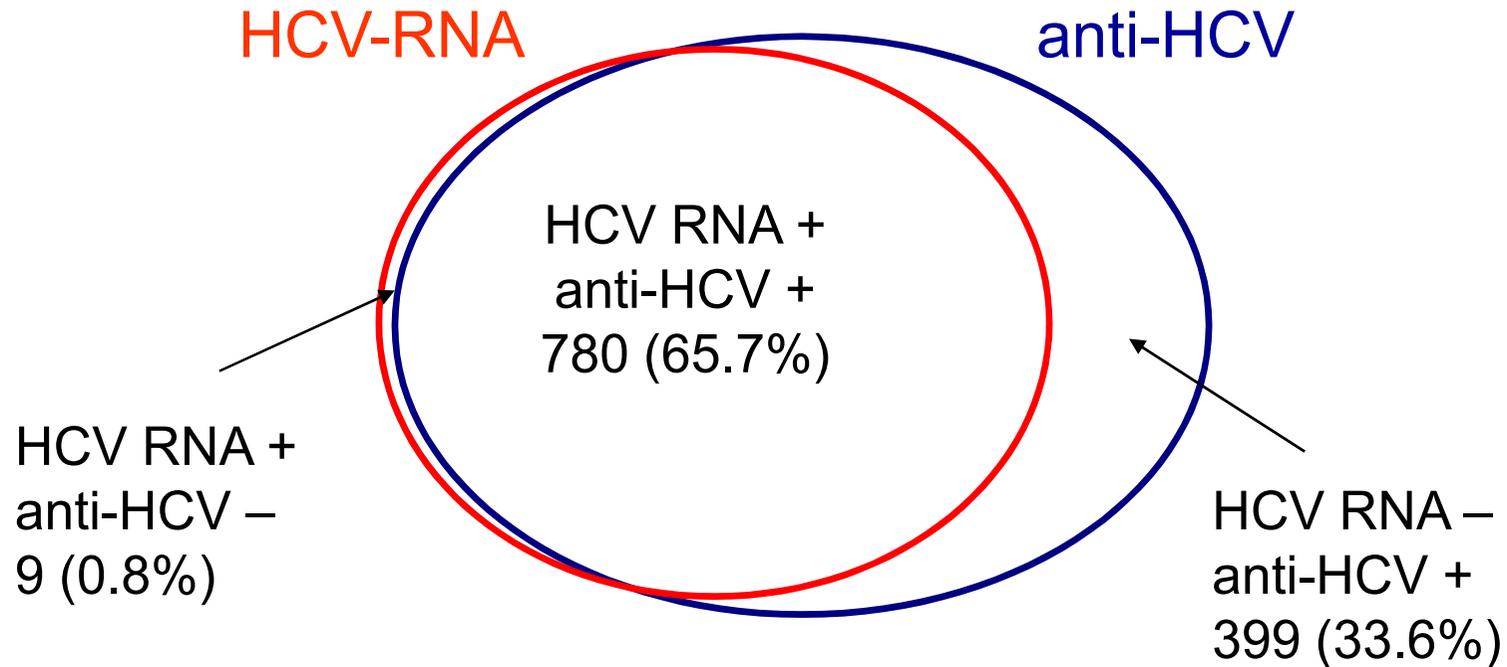
- in pre-seroconversion phase: 3 (1-10 virions)^{1,2,3,4}
- in HB vaccine breakthrough infection (?)
- in clearance or early recovery phase (?)
- in chronic viremia phase: 300 (100-1000) virions^{1,5,6,7}
- in chronic occult HBV infection phase: (?)
- in HBsAg positive low viral load carriers: (?)
- in HIV elite controllers: (?)
- in HCV low viral load (occult) carriers: (?)

(?) proposed to assume ID_{50} of 300 (100-1000) virions in plasma in modelling

1) Ma ZM et al, J of Virology, 2009, 83:3288-97 2) Komiya K et al. Transfusion 2008;48:286 3). Katayama K et al, Intervirology, 2004, 47, 57 4) Kleinman S et al Vox Sang 96, Suppl1, ISBT abstract 5). Tabuchi A et al. J Med Virol 2008;80:2064-8 6) Hijikata J. Virol 1993 67:1953 7) Alter H et al. J. Viral Hep 1995, 2:121;

Preliminary HCV and HIV data

HCV screening results of first time donations in Egypt

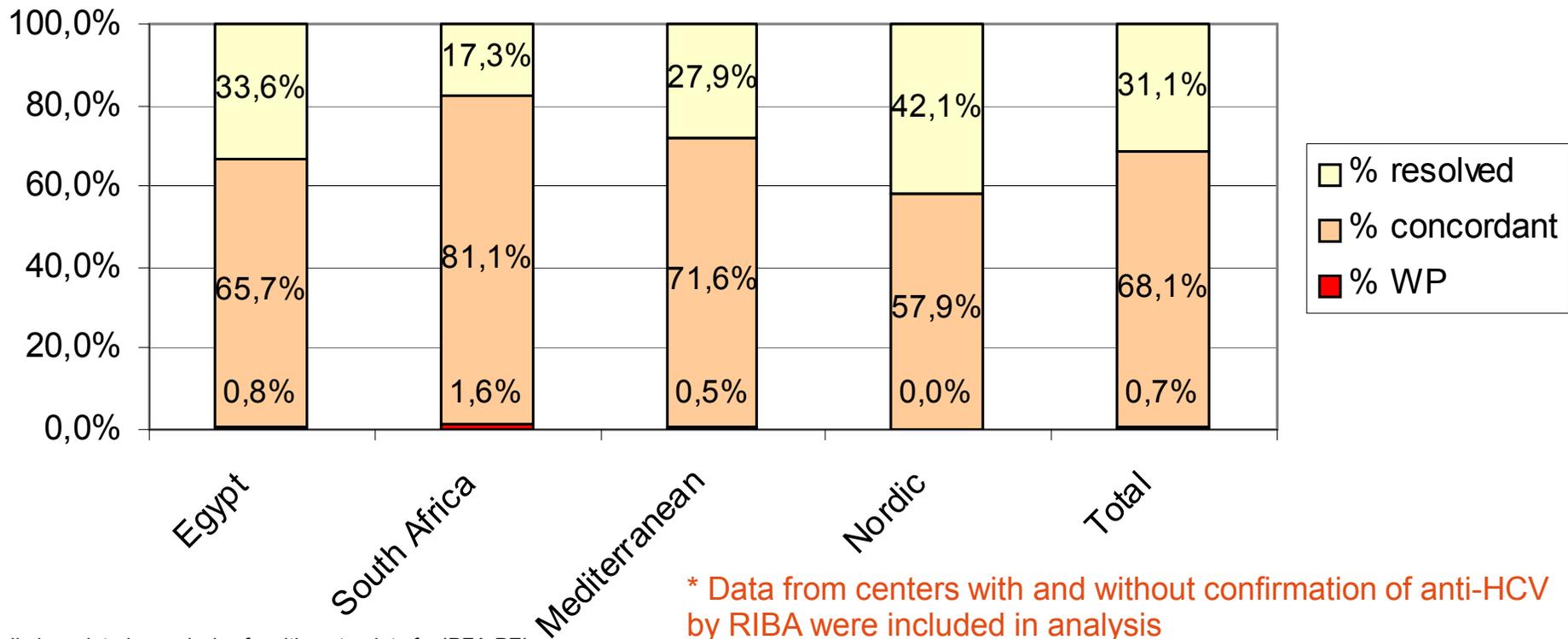


1188 HCV infections in 27,655 first time donors
(prevalence 4.3%)

El Ekiaby M et al. and Goubran et al. Vox Sang 96, Suppl1, ISBT abstracts, Cairo, March, 2009

Proportion HCV-RNA and antibody positives* in HCV infected first time donors from 4 regions

	Egypt	South-Africa	Mediterranean	Nordic	Total
First time donors	27,655	327,968	231,221	43,058	629,902
Infections (prevalence)	1188 (4.3%)	148 (0.05%)	419 (0.18%)	19 (0.04%)	1753 (0.28%)
WP NAT yield (rate)	9 (1:3070)	2 (1:164,000)	2 (1:116,000)	0 (0.00%)	13 (1:48,454)

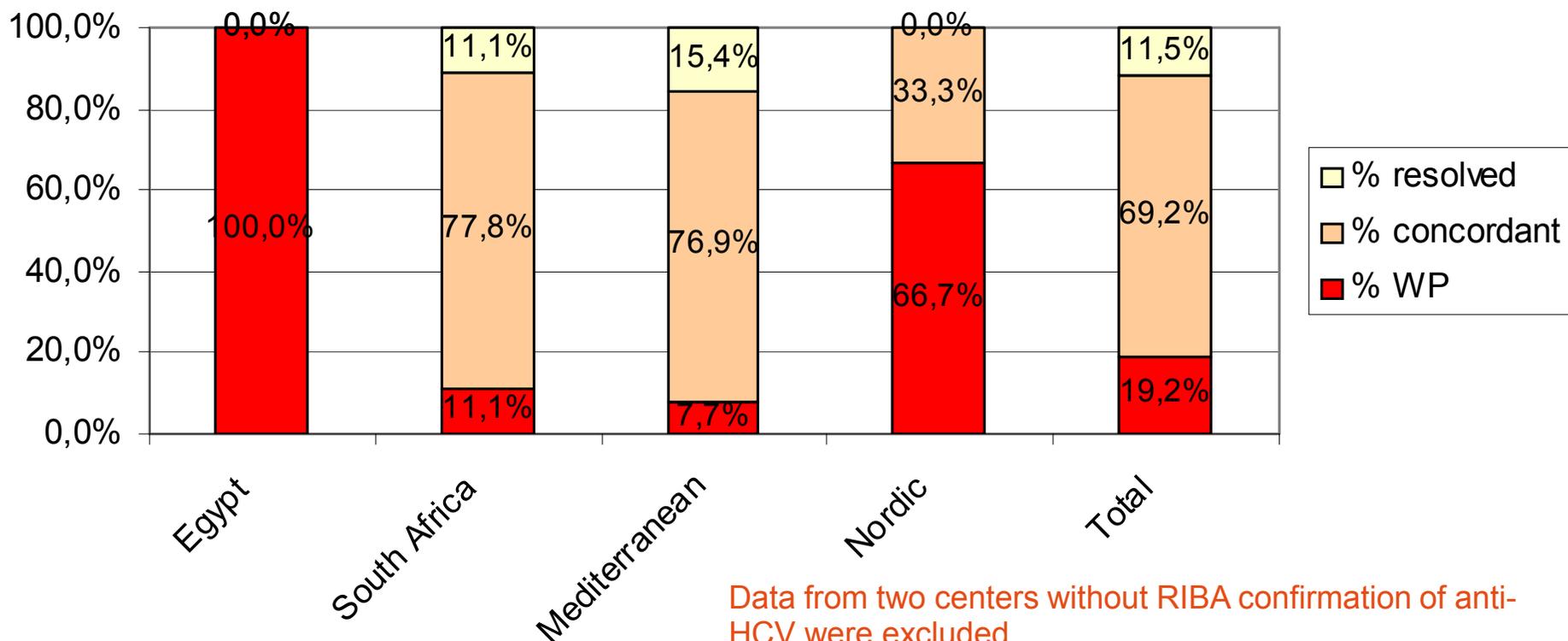


* Data from centers with and without confirmation of anti-HCV by RIBA were included in analysis

Proportion HCV-RNA and antibody positives in acutely infected repeat donors from 4 regions

	Egypt	South-Africa	Mediterranean	Nordic	Total
Repeat donors	12,644	1,769,441	1,431,726	507,750	3,721,561
infections (rate)	1 (1:12,644)	9 (1:196,605)	13 (1:110,133)	3 (1:169,250)	26 (1:143,137)
WP NAT yield (rate)	1 (1:12,644)	1 (1:1,769,441)	1 (1:1,431,726)	2 (1:253,875)	5 (1:744,312)

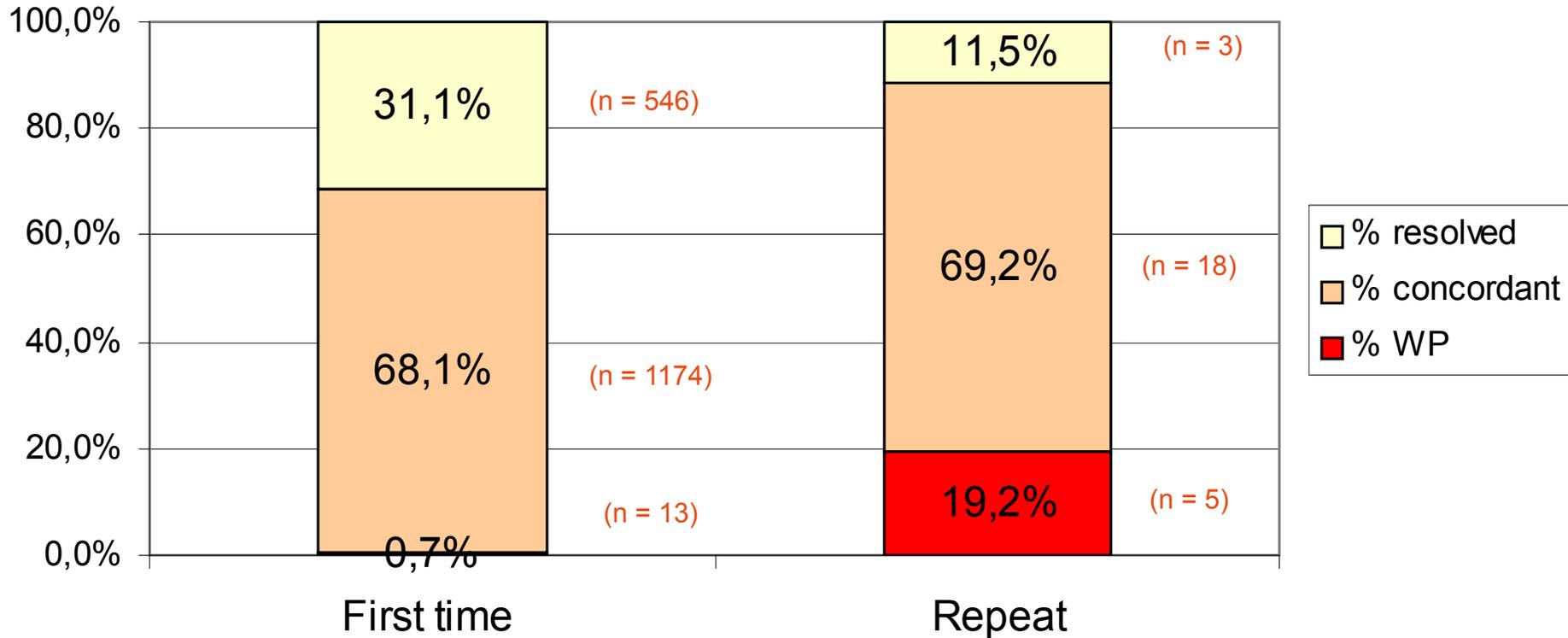
HCV infections in lapsed donations (interval >365 days) excluded



Data from two centers without RIBA confirmation of anti-HCV were excluded

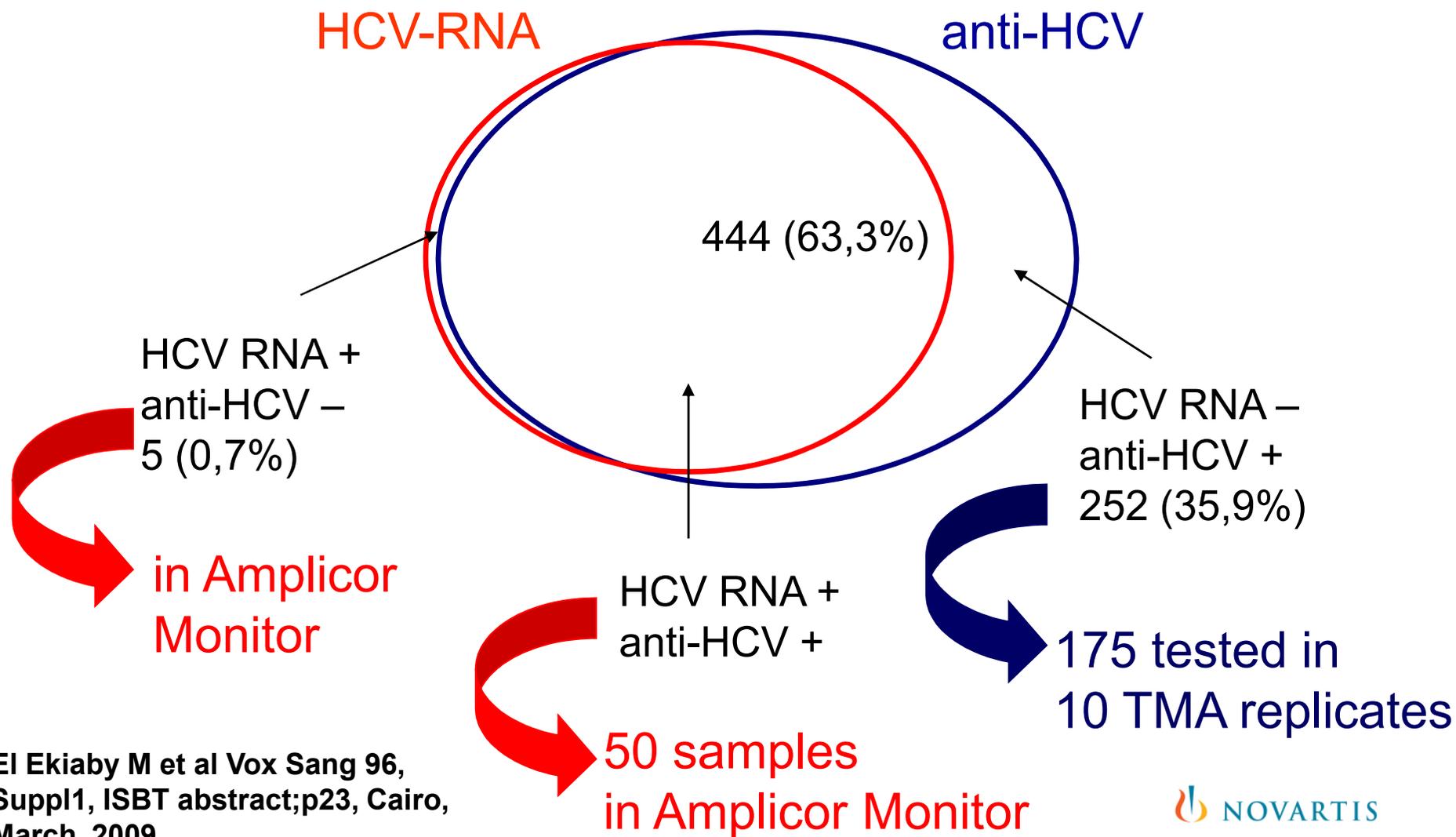
Proportion HCV-RNA and antibody positives in first time and repeat donations (interval <365 days)

	First time	Repeat
donations	629,902	3,721,561
HCV infections	1774	26



Data from two centers were excluded from repeat donation data set because anti-HCV reactivity was not confirmed by RIBA. Also infections in lapsed donations with intervals >365 days were excluded from repeat donation data.

Viral load analysis



El Ekiaby M et al Vox Sang 96, Suppl1, ISBT abstract;p23, Cairo, March, 2009

Replicate TMA results in subset of anti-HCV+/RNA- samples

- Random subset of 175 out of 252 anti-HCV+/RNA- samples were tested in 10 replicates
- 173/175 (98,8%) were non reactive in 10 replicates
- One donation was reactive in 3/25 (12%) of replicates and contained ~0.5 copies/ml plasma*
- Another donation was reactive in 10/25 (40%) of replicates and contained ~1.8 copies/ml plasma*

*determined by probit analysis against replicate TMA results on HCV genotype 1 and 4 standard dilutions

Viral load in 5 HCV window period samples in Egypt (NAT yield rate 1:3,100)

donor	HCV-RNA copies/ml
1	1.7E+07
2	2.0E+07
3	3.2E+06
4	9.1E+05
5	82*

dilution	TMA reactive*
1:1	24/24 (100%)
1:4	22/24 (92%)
1:8	14/24 (58%)
1:16	12/24 (50%)
1:32	9/24 (38%)
1:64	1/24 (4%)

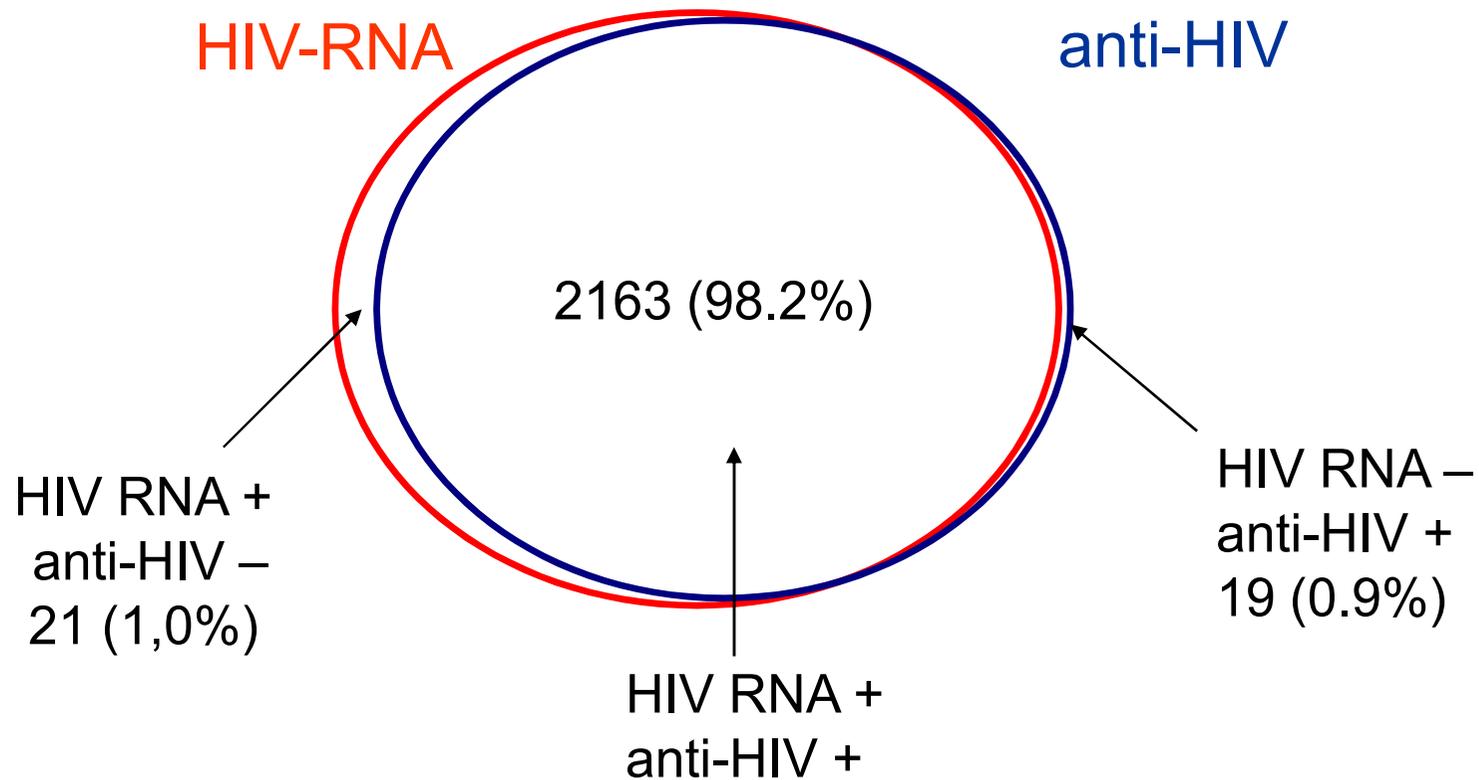
* determined by probit analysis against replicate TMA results on HCV genotype 1 and 4 standard dilutions

Modeling residual risk of HCV transmission by RBCs from Egyptian first time donors with different screening scenarios

Screening with:	WP risk by transfusion of RBCs ^{1,2}	
	risk day equivalents	Risk
Anti-HCV only	65	1:3100
HCV Ag/Ab Combo only	39 ³	1:5200 ¹
ID NAT only	1.5 ⁴	1:135,700
Serology + ID NAT	1.3	1:151,900
Serology + MP 16 NAT	3.1	1:65,600

1. adapted from WP ratio model (Busch MP et al. Transfusion 2005;45:254-264)
2. Transmission risk model (Weusten J et al, Transfusion 2002;42:537-548).
3. 60% of WP samples missed by Biorad combo ELISA
4. assumed both ramp up and recovery WPs

HIV infections in first time donors during three years of ID-NAT screening in South Africa (SANBS, WPBTS)

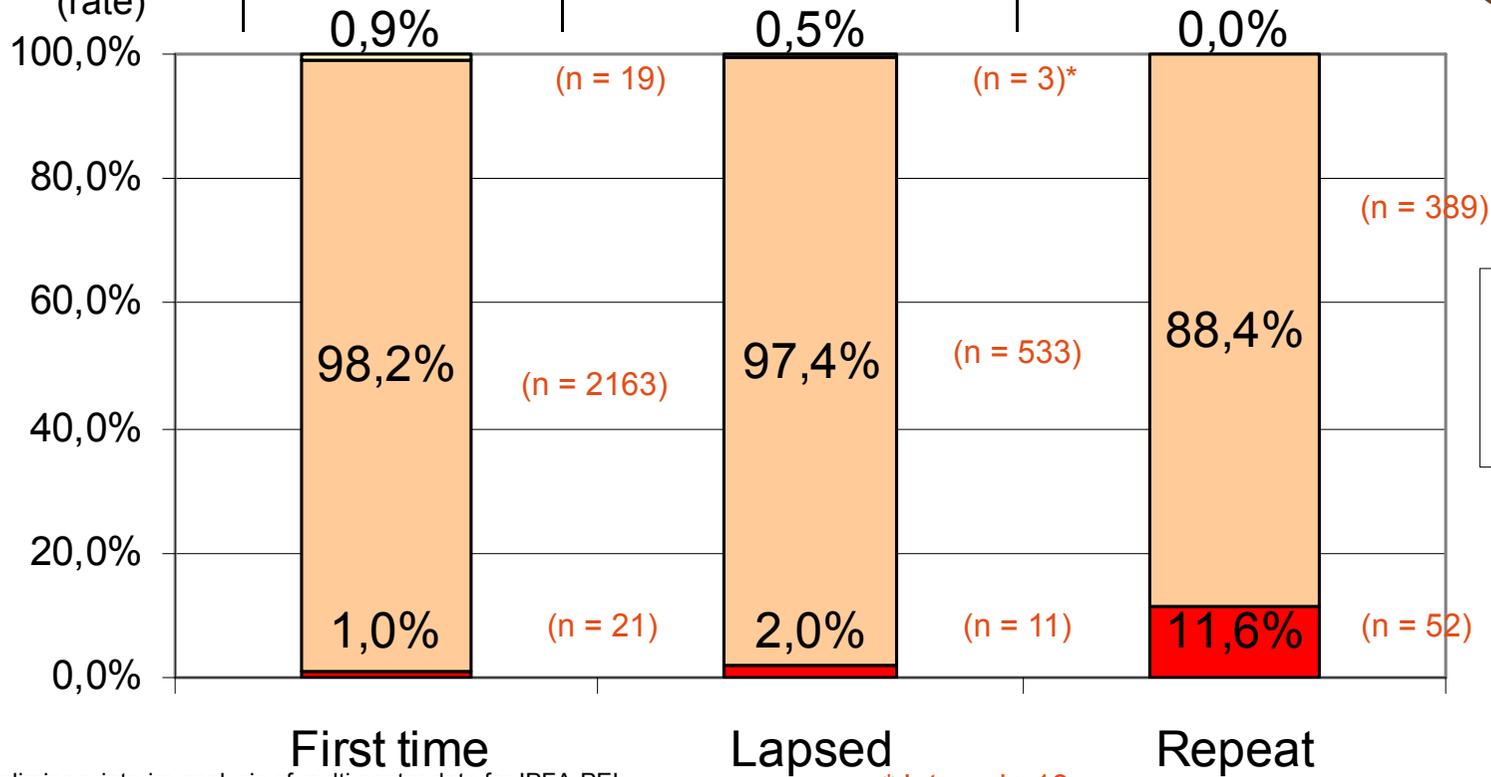


2203 confirmed HIV infections in 327,928 first time donations

Proportion detectable HIV-RNA in first time, lapsed, and repeat donors in South Africa (SANBS, WPBTS)

	First time	Lapsed	Repeat
donations	327,968	125,609	2,087,402
infections (prevalence)	2203 (0.67%)	547 (0.44%)	450 (0.02%)
NAT WP yield (rate)	21 (1:15,618)	11 (1:11,419)	52 (1:40,142)

2.51 and 3.51 fold lower yield rate than in first time and lapsed donors



Preliminary interim analysis of multi-center data for IPFA-PEI meeting in Zagreb by Kleinman et al

* interval >13 yr

HIV infections in first time donations from three regions

Region	South-Africa	Mediterranean	Nordic
donations	327,968	231,261	43,058
WP yield	21	1	0
All infections	2203	51	1
prevalence	1:149	1:4535	1:43,058
NAT yield rate	1:15,618	1:231,261	
% WP	1.0%	2.0%	0%
% concordant	98.2%	98.0%	100%
% elite controller	0.9%	0%	0%

HIV infections in repeat donations from three regions

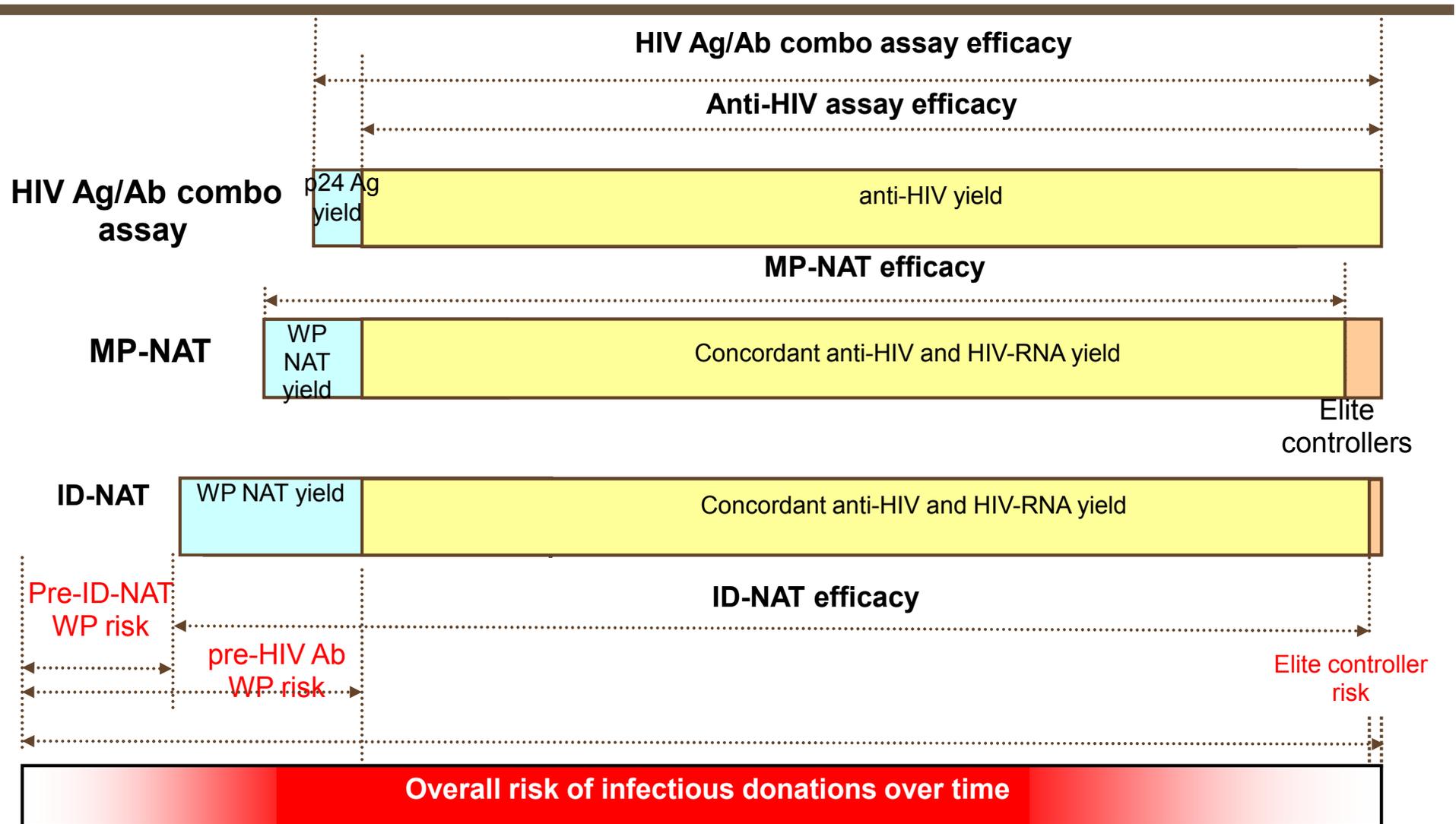
Region	South-Africa	Mediterranean	Nordic
donations	2,087,402	1,851,220	507,750
WP yield	52	4	1
All infections	450	71	2
Seroconv. rate	1:4639	1:26,074	1:253,875
NAT yield rate	1:40,142	1:462,805	1:507,750
% WP	11.6%	5.6%	50%
% concordant	88.4%	94.4%	50%
% elite controller	0%	0%	0%

Estimated viral load in 21 HIV elite controllers*(SANBS, 3 year)

Year 1					Year 2				
unit nr	pos	N test	% pos	cps/ml	unit nr	pos	N test	% pos	cps/ml
1	6	30	20%	0.3	3	destroyed			
2	9	30	30%	0.4	4	8	20	40%	0.6
Year 3					5	12	25	48%	0.8
14	destroyed				6	2	20	10%	0.2
15	4	10	40%	0.6	7	0	31	0%	<0.1
16	6	6	100%	2.7	8	20	23	87%	2.9
17	0	20	0%	<0.1	9	2	30	7%	0.1
18	12	20	60%	1.1	10	4	25	16%	0.3
19	1	29	3%	0.1	11	0	10	0%	<0.2
20	2	20	10%	0.2	12	0	33	0%	<0.1
21	4	25	16%	0.3	13	5	25	20%	0.3

*determined by probit analysis against HIV subtype C standard calibrated in bDNA copies/ml
(Vermeulen et al, to be published)

Efficacy of HIV screening assays in repeat or lapsed donations



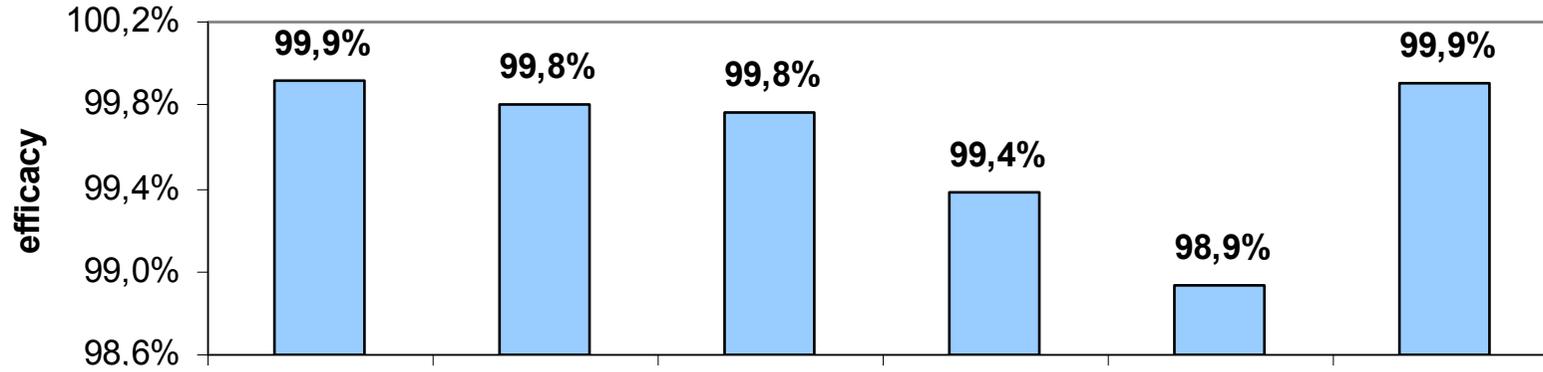
HIV transmission risk* per million donations in different screening scenarios (SANBS 3 years)

screening scenario	risk per million			
	First time	Lapsed	Repeat	All
ID-NAT + anti-HIV	5,63	9,72	2,26	3,02
MP-8 NAT + anti-HIV	13,50	23,31	5,42	7,24
MP16-NAT + anti-HIV	16,12	27,83	6,47	8,64
p24 Ag + anti-HIV	43,10	74,41	17,30	23,10
anti-HIV only	74,62	129,03	29,95	39,99
ID-NAT only	6,22	9,72	2,26	3,10

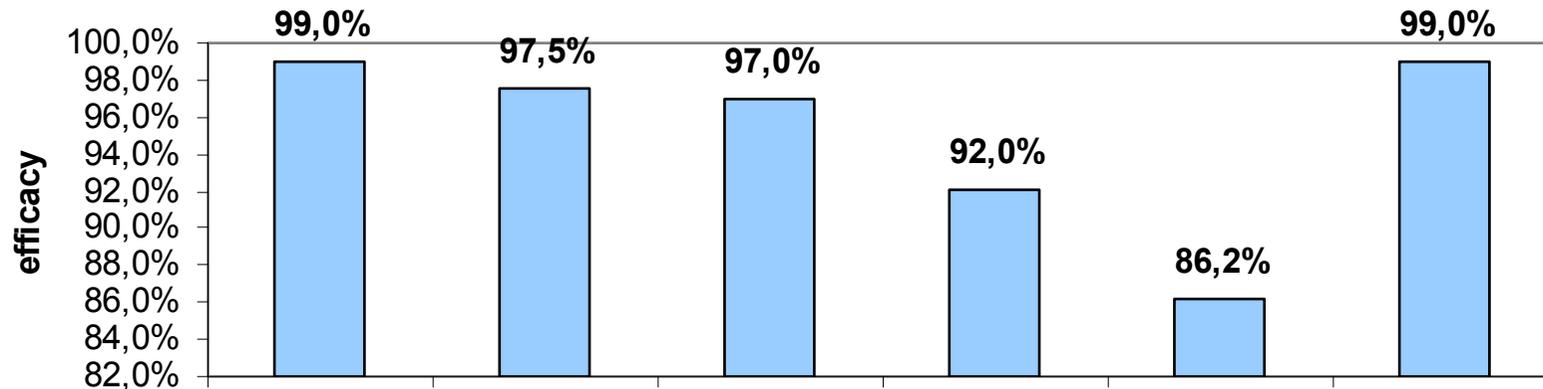
*assuming ID₅₀ of 3.2 virions for HIV-NAT WP and 320 virions in elite controllers

HIV Screening Efficacy (SANBS 3 years)

First time donations



repeat donations



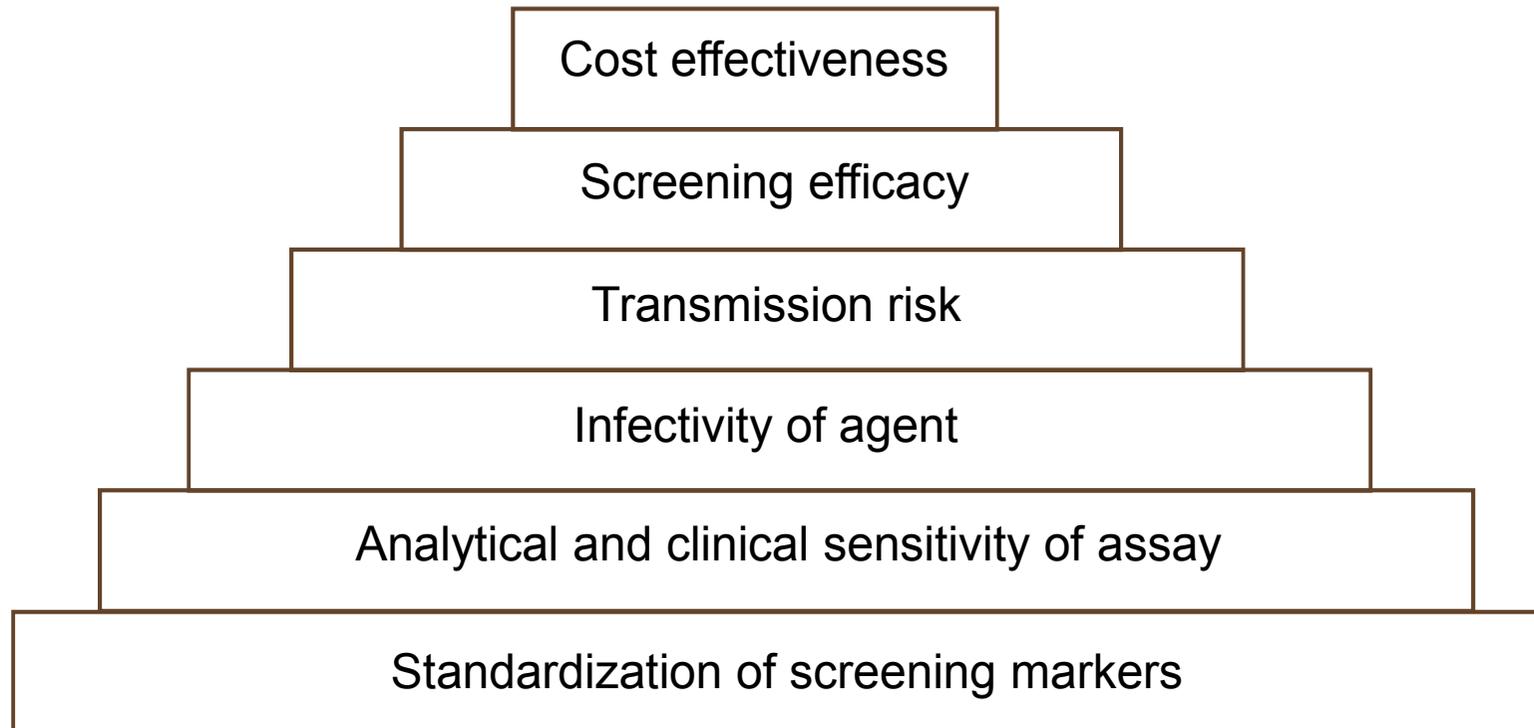
ID-NAT + anti-HIV MP-8 NAT + anti-HIV MP16-NAT + anti-HIV p24 Ag + anti-HIV anti-HIV only ID-NAT only

screening scenario

Next steps

- Obtain data from additional countries and use these to extend the HIV and HCV risk analyses
- Perform similar analyses for HBV
- Possibly extend the protocol to include data collection from countries using other NAT testing:
 - MP NAT with Ultrio
 - MP NAT with MPX/s201 or Ultrio Plus
 - ID NAT with Ultrio Plus
- Conduct cost-effectiveness analyses

Foundations for a robust analysis of screening efficacy and cost-effectiveness



Co-investigators and laboratories who have supplied data

- Marion Vermeulen, Ravi Reddy, South African National Blood Service, Johannesburg, South Africa
- Arthur Bird, Russell Cable, Western Province Blood Transfusion Service, Cape Town, South Africa
- Heidi Goubran, Faten Mofteh, National Blood Transfusion Service, Cairo, Egypt
- Magdy El Ekiaby, Shabrawishi Hospital, Dokki, Egypt
- Paola Ghiazza, St Anna Hospital, Turin, Italy
- Paola Manzini University of Turin, Turin, Italy
- Roberto Roig, Valencia Regional Blood Transfusion Center, Valencia, Spain
- Sylvia Sauleda, Banc de Sang I Teixits, Barcelona, Spain
- Christoph Niederhauser, Martin Stolz, Blood Transfusion Service SRC Berne, Berne, Switzerland
- Snezna Levicnik, Blood Transfusion Center of Slovenia, Ljubljana, Slovenia
- Sussanne Wessberg, Sussane Elkblom, Mervi Lankinen, Finnish Red Cross Blood Service, Helsinki, Finland,
- Peter Flanagan, New Zealand Blood Service, Auckland, New Zealand
- Tsoi Wai Chiu, Kit Che Lin, Hong Kong Red Cross Blood Center, Hong Kong

Co-investigators and laboratories who have agreed to supply data

- Michelina Miceli, Forlanini Hospital, Rome, Italy
- Pilar Torres, Community Blood Center, Madrid, Spain
- Rocio Gonzalez, Emma Castro, Red Cross Blood Center, Madrid, Spain
- Henrik Ulm, Lene Harritshoj, Copenhagen University Hospital, Copenhagen, Denmark
- Ewa Brojer, Piotr Crabarzyk, Institute of Haematology and Transfusion Medicine, Warsaw, Poland
- Jolanta Gdowska, Dariusz Piotrowski, Warsaw Blood Center, Warsaw, Poland
- Sally Lam, Diane Teo, Health Systems Agency, Singapore
- Abdul Hamid, Malaysia Ministry of Health Blood Transfusion Organization